Halifax Road, Penistone



Project:	Halifax Road, Penistone	Site Inspection reference:	SI-4578-01			
Date of inspection:	20.05.2020	No. of Pages:	5			
Inspection carried out by:	Olivia Benson BSc (Hons) Graduate Ecologist					
Task description:	<u>eDNA survey</u> Two offsite ponds, referred to as Ponds 2 & 3 in Brooks Ecological's Preliminary Ecological Appraisal Report ER-3706-01-B, were surveyed to confirm the presence or likely absence of great crested newt (GCN). These ponds are associated with fishing, with access being provided by Alan Jackson, the ponds groundskeeper via Steven Green at Yorkshire Land Ltd, the site landowner. The eDNA survey involved taking water samples from 20 different locations around the pond's margin, focusing on areas where newts are most likely to gather. The survey used sterile kits supplied by Surescreen Scientifics laboratory and followed methodology as advised in the Natural England Technical Advice Note (WC1067).					
Results:	Water samples were collected on 20 th May 2020 and the results were returned on 27 th May 2020 - see Appendix 1 for full report. A negative result was returned for both ponds, meaning that GCN DNA was <u>not</u> detected within the water sampled collected. This result can be used to demonstrate the likely absence of GCN from these ponds.					
Conclusions and Recommendations:	Based on this information present on the site is ne mitigation is considered n	, it is concluded that the gligible and no furthe ecessary.	ne risk of GCN being er survey or specific			







Folio No:	E7454
Report No:	1
Purchase Order:	4578a
Client:	BROOKS ECOLOGICAL
Contact:	Olivia Benson

TECHNICAL REPORT

ANALYSIS OF ENVIRONMENTAL DNA IN POND WATER FOR THE DETECTION OF GREAT **CRESTED NEWTS (TRITURUS CRISTATUS)**

SUMMARY

When great crested newts (GCN), Triturus cristatus, inhabit a pond, they continuously release small amounts of their DNA into the environment. By collecting and analysing water samples, we can detect these small traces of environmental DNA (eDNA) to confirm GCN habitation or establish GCN absence.

RESULTS

Date sample received at Laboratory: Date Reported: Matters Affecting Results:			y:	21/05/ 27/05/ None	2020 2020			
Lab Sample No.	Site Name	O/S Reference	SI	IC	DC	IC	Result	Positive Replicates
0503	Scout Dike		Pa	ass	Pass	Pass	Negative	0

If you have any questions regarding results, please contact us: ForensicEcology@surescreen.com

Reported by: Chris Troth

Approved by: Sarah Evans



Forensic Scientists and Consultant Engineers SureScreen Scientifics Ltd, Morley Retreat, Church Lane, Morley, Derbyshire, DE7 6DE UK Tel: +44 (0)1332 292003 Email: scientifics@surescreen.com Company Registration No. 08950940

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METHODOLOGY

The samples detailed above have been analysed for the presence of GCN eDNA following the protocol stated in DEFRA WC1067 'Analytical and methodological development for improved surveillance of the Great Crested Newt, Appendix 5.' (Biggs et al. 2014). Each of the 6 sub-sample tubes are first centrifuged and pooled together into a single sample which then undergoes DNA extraction. The extracted sample is then analysed using real time PCR (qPCR), which uses species-specific molecular markers to amplify GCN DNA within a sample. These markers are unique to GCN DNA, meaning that there should be no detection of closely related species.

If GCN DNA is present, the DNA is amplified up to a detectable level, resulting in positive species detection. If GCN DNA is not present then amplification does not occur, and a negative result is recorded.

Analysis of eDNA requires scrupulous attention to detail to prevent risk of contamination. True positive controls, negative controls and spiked synthetic DNA are included in every analysis and these have to be correct before any result is declared and reported. Stages of the DNA analysis are also conducted in different buildings at our premises for added security.

SureScreen Scientifics Ltd is ISO9001 accredited and participate in Natural England's proficiency testing scheme for GCN eDNA testing. We also carry out regular inter-laboratory checks on accuracy of results as part of our quality control procedures.

INTERPRETATION OF RESULTS

SIC:	Sample Integrity Check [Pass/Fail] When samples are received in the laboratory, they are inspected for any tube leakage, suitability of sample (not too much mud or weed etc.) and absence of any factors that could potentially lead to inconclusive results.
DC:	Degradation Check [Pass/Fail] Analysis of the spiked DNA marker to see if there has been degradation of the kit or sample between the date it was made to the date of analysis. Degradation of the spiked DNA marker may lead indicate a risk of false negative results.
IC:	Inhibition Check [Pass/Fail] The presence of inhibitors within a sample are assessed using a DNA marker. If inhibition is detected, samples are purified and re-analysed. Inhibitors cannot always be removed, if the inhibition check fails, the sample should be re-collected.
Result:	 Presence of GCN eDNA [Positive/Negative/Inconclusive] Positive: GCN DNA was identified within the sample, indicative of GCN presence within the sampling location at the time the sample was taken or within the recent past at the sampling location. Positive Replicates: Number of positive qPCR replicates out of a series of 12. If one or more of these are found to be positive the pond is declared positive for GCN presence. It may be assumed that small fractions of positive analyses suggest low level presence, but this cannot currently be used for population studies. In accordance with Natural England protocol, even a score of 1/12 is declared positive. 0/12 indicates negative GCN presence. Negative: GCN eDNA was not detected or is below the threshold detection level and the test result should be considered as evidence of GCN absence, however, does not exclude the potential for GCN presence below the limit of detection.





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0495	Fishing Pond	SE 244 041	Pass		Pass	Pass	Negative		0	

If you have any questions regarding results, please contact us: ForensicEcology@surescreen.com

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